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Human transferrin and lactoferrin cooperatively support colonization by *Neisseria meningitidis* in the murine nasopharynx

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### Background

*Neisseria meningitidis* (Nm) is a regular colonizer of the human nasopharynx, whose human-restricted nature makes in vivo studies of host-pathogen interaction difficult. Expression of human CEACAM1 (hCCM1) is required for Nm colonization of mouse nasal passages. Nm also has an exquisite specificity for the human forms of transferrin (hTf) and lactoferrin (hLf), being unable to utilize iron from mouse homologues of these proteins when infecting the murine nasopharynx.

### Aim/Methods

To overcome this host restriction, we have used transgenic mice expressing various combinations of hCCM1, hTf and/or hLf to uncover the role of iron sources during Nm colonization.

### Results

In vitro studies revealed that serum from hTf- or hLf-transgenic mice supported Nm growth, but serum from their wild type littermates did not. Notably, growth in the transgenic mice serum was comparable to that seen in human sera. Nasal infection of hCCM1 mice with iron-starved Nm resulted in no recoverable meningococci

at 7 days post-infection. Mice co-expressing hTf and/or hLf along with hCCM1 allowed for Nm recovery at both 7 and 10 days post-infection. Notably, the expression of all 3 transgenes resulted in the highest median burden and colonization rates, suggesting that the two iron sources may have different contributions to Nm infection. The contribution of the two bacterial receptor systems was further explored by a competitive infection with a mixture of  $\Delta$ tbpB and  $\Delta$ lbpB strains of Nm in the triple transgenic (hCCM1/hTf/hLf) mice. At 3 days post-infection, 75% of mice were colonized by higher proportion of  $\Delta$ lbpB Nm, suggesting that transferrin may be more useful during early colonization.

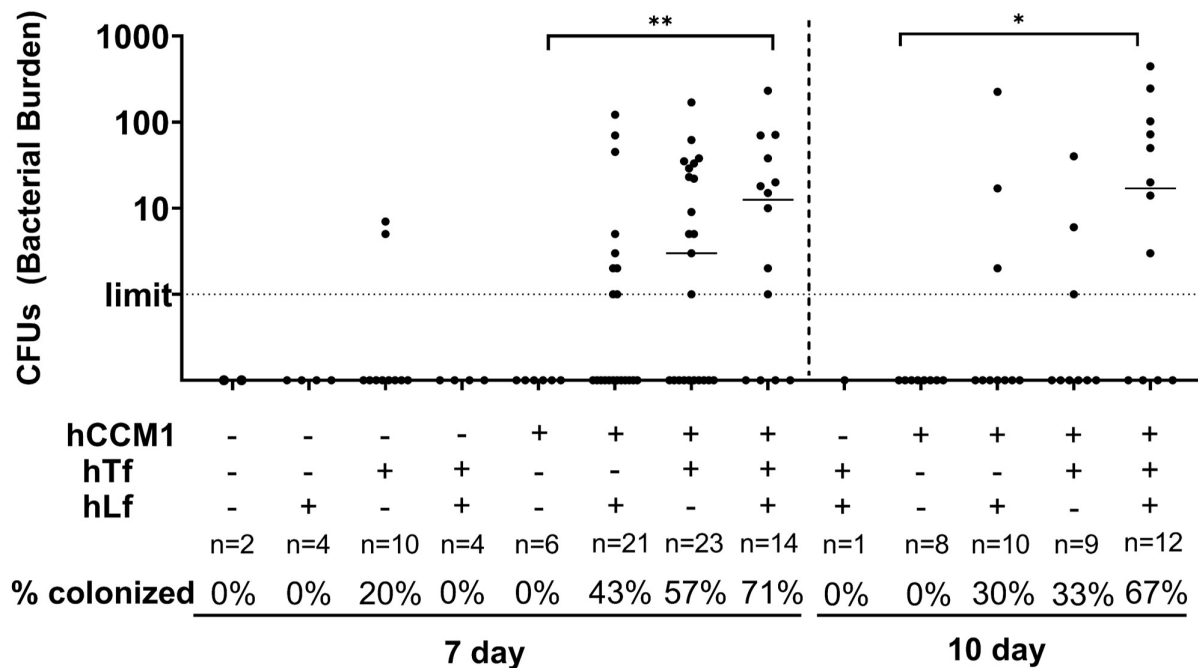
## Conclusions

Collectively, this work establishes critical contributions of the meningococcal transferrin and lactoferrin receptors during nasal colonization and provides a novel in vivo model to better study Nm host-pathogen interaction at the mucosa.

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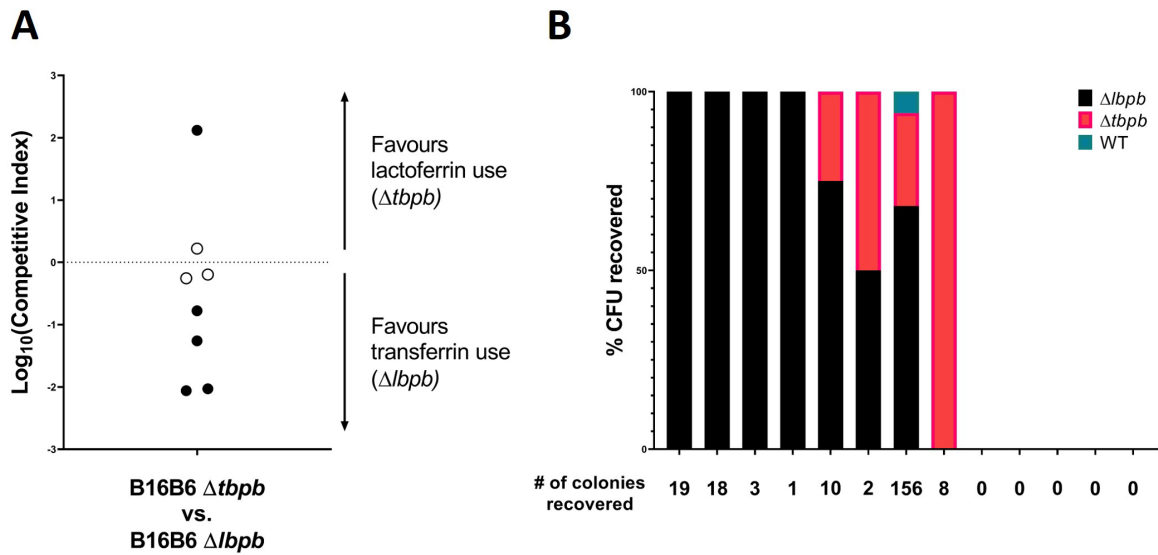
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[Combined expression of hTf and hLf with hCEACAM1 enhances nasopharyngeal colonization rates at day 7 and 10 post-infection](#)



**Supplemental Figure 1. Combined expression of hTf and hLf with hCEACAM1 enhances nasopharyngeal colonization rates at day 7 and 10 post-infection.** FvB mice were nasally infected with  $10^6$  CFU/mouse of *Nme* B16B6. Bacterial enumeration was done 7-and 10-days post-infection via nasal swabs. Data are pooled from 3 independent experiments. Limit represents limit of detection; bars represent group median. Kruskal-Wallis test with multiple comparisons. \*\* $p < .01$ , \* $p < .05$ . hCCM1, human CEACAM1; hTF, human transferrin; hLF, human lactoferrin.

Utilization of hTf may be more advantageous than hLf during early *N. meningitidis* colonization of the murine nasopharynx



**Supplemental Figure 2. Utilization of hTf may be more advantageous than hLf during early *N. meningitidis* colonization of the murine nasopharynx.** FvB hCEACAM1<sup>+</sup> hTf<sup>+</sup> hLf<sup>+</sup> mice were nasally infected with a 1:1 mix of *Nme* B16B6 deficient in TbpB ( $\Delta tpb$ ) or LbpB ( $\Delta lbp$ ). Bacterial enumeration was done 5-days post-infection via nasal swabs. **A)** Competitive index of  $\Delta tpb$  vs.  $\Delta lbp$  *Nme*. **B)** Percentage of each mutant colonies recovered per mouse. Competitive index = ( $\Delta tpb$ : $\Delta lbp$ )/ ( $\Delta tpb$  inoculum concentration/ $\Delta lbp$  inoculum concentration). Log<sub>10</sub>[Competitive index] > 0 = selective advantage for  $\Delta tpb$ ; Log<sub>10</sub>[Competitive index] < 0 = selective advantage for  $\Delta lbp$ . For the purpose of the calculation, value of 0.1 was used when no colonies were recovered for either mutant. Empty circles in **A)** indicate mice with mixed population of knockout strains recovered.